The Seleno-acetal Route to 1α -Hydroxy-vitamin D Analogues: Synthesis of 24-Oxa- 1α -hydroxy-vitamin D₃, A Useful Vitamin D Metabolism Probe

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Abstract. Alkylation of the lithio-demethylseleno-derivative of seleno-acetal 11 with chloromethyl isopropyl ether [shown by NMR to give the (22S)-methylseleno-compound 13a as the major diastereoisomer] is the key reaction in the synthesis of the 24-oxa analogue (MC 1090, 8) of 1α -hydroxyvitamin D₃ (4) The metabolism of 8 to calcitroic acid (7) is demonstrated *in vitro* in a hepatocyte cell (Hep 3B) model This supports the hypothesis that 8 can undergo enzymatic 25-hydroxylation analogous to the activation of 4, or a similar side chain hydroxylation, but then short cuts the target cell side chain cleavage pathway taking 1α ,25-dihydroxyvitamin D₃ to 7.

Vitamin D₃ (1), formed naturally in the skin by (photo- followed by thermal-) isomerisation of 7dehydrocholesterol, is transported bound to a protein in the plasma to the liver where 25-hydroxylation occurs, the first step of its metabolic activation required for function Further metabolism of 25hydroxyvitamin D₃ (25-OH-D₃, 2) to the hormonally active 1α ,25-dihydroxyvitamin D₃ (1,25-(OH)₂-D₃, 3) takes place in the kidney and is under strict feedback regulation. The interaction of 1,25-(OH)₂-D₃ with a specific receptor present in the classical target cells (intestine, bone, kidney) initiates a cascade of events resulting in the transport of calcium ions necessary for homeostasis ¹ Target cell catabolism of 1,25-(OH)₂-D₃ (3) by 1,25-(OH)₂-D₃ -inducible enzymes is believed to proceed sequentially *via* the 24(*R*)-hydroxy-, 24-oxo-, and 23(*S*)-hydroxy-24-oxo-derivatives to the inactive, side chain cleaved C₂₃ compound calcitroic



X = Y = H	Vitamin D ₃
X = H, Y = OH	25-OH-D ₃
X = Y = OH	1,25-(OH) ₂ -D ₃
X = OH, Y = H	1α-OH-D ₃





Scheme 1 Target cell catabolism of 1,25-(OH)₂-D₃ (3)

acid (7) (Scheme 1), which is transported to the liver and excreted in the bile ² The C₂₃ alcohol (6) has also been identified on this pathway and suggested to be the precursor of calcitroic acid ³ Our experiments with a synthetic sample of the putative pivotal intermediate C₂₃ aldehyde (5) indicate that the alcohol is probably a side product of the pathway, though both can be converted by a 1,25-(OH)₂-D₃-inducible enzyme to calcitroic acid ⁴

The liver vitamin-D₃-25-hydroxylase has been shown to possess some variability in its substrate selectivity Thus, the synthetic analogue 1α -hydroxyvitamin D₃ (1α -OH-D₃, 4) is hepatically activated to 1,25-(OH)₂-D₃ (3) (without feedback control) and is a therapeutically important prodrug for the hormone ⁵ A modified side chain can also be hydroxylated, as is the case with vitamin D₂ ⁶ A human hepatocyte (Hep 3B) cell model has been shown to provide an accurate *in vitro* reflection of the *in vivo* 25-hydroxylation of vitamin D₃⁷ and analogues (including 4) and has also provided an insight into the possibility of varying the hydroxylation site by structural modification. For example, the 26,27-cyclo-derivative of 1α -OH-D₃ (MC 969) is exclusively 24-hydroxylated by this system ⁸

To probe further the details of vitamin D catabolism we have designed an analogue, the 24-oxaderivative of 1α -OH-D₃ (8, MC 1090 = 24-oxa-4), which, if a substrate for the vitamin-D-hydroxylase, can short cut the calcitroic acid pathway Thus, 25-hydroxylation of 8 would give the C₂₃ alcohol (6) directly from decomposition of the unstable hemi-acetal intermediate (9) (Scheme 2) An alternative possibility, 23-hydroxylation to 10, would give the C₂₃ aldehyde (5) Either of these side chain cleaved compounds could then be converted to 7 We report here a synthesis of MC 1090 together with the results of the *in vitro* metabolism studies ⁹ The metabolism studies have been published in abstract form.¹⁰



Scheme 2 Proposed metabolic side chain cleavage pathways for MC 1090 (8)

Alkylation of the lithio-demethylseleno-derivative (12) of seleno-acetal 11 to give two series of 22methylseleno-compounds (cf. Scheme 3) constitutes the key reaction in the synthesis of a variety of side chain modified analogues of $1,25-(OH)_2-D_3$ (3),¹¹⁻¹⁵ including 1α -OH-D₃ (4), for which the alkylating agent (R-Hal) is isoamyl bromide ¹¹ A corresponding alkylation with chloromethyl isopropyl ether (as illustrated) gave the anticipated intermediates 13 for the MC 1090 synthesis, though in this case the diastereoisomers at C-22 were chromatographically inseparable, and recrystallisation only marginally enriched the mixture, which therefore had to be used in the subsequent steps as such. The kinetic ratio of the 22-epimers in 13 was readily determined as 3.2 from the NMR-spectra (see Experimental Section). A modest selectivity in this reductive alkylation of 11 was expected on the basis of our experience with alkylations using unactivated primary alkyl halides¹¹ and also a reductive formylation¹³ and hydroxy-alkylation¹⁴ of 11, for



Scheme 3. * R-Hal = Cl-CH₂OCH(CH₃)₂, # R = -CH₂OCH(CH₃)₂



which reactions we were able to demonstrate the preferential formal substitution of the pro-R selenomethyl group. For the conversion of 11 to 13 we can convincingly assign the same stereoselectivity by NMR comparison (see Table) of the product mixture with the previously described¹³ alcohols 14, whose configurations have been deduced. The juxtaposition of data shown clearly allows correlation of 22S-14 with the major isomer 13 (thus assigned the structure 13a) and of 22R-14 with the minor isomer (viz. 13b).¹⁶

Table. Chemical Shifts⁴ for Selected Signals in the NMR Spectra of Compounds 13 and 14.^b

	<i>C</i> -18	C-21	C-20	C-23	SeCH ₃
13	11 5 / 12.2	15 6 / 14.6	40.3 / 36 4	69.1 / 70.8	4.5 / 4.1
Δδ°	-0.7	1.0	3.9	-1.7	0.4
(22 <i>S/R</i>)-14	11.5 / 12 2	16 1 / 15.3	41 1 / 37 6	61 2 / 64.8	4.8 / 4.4
Δδ ^d	-0.7	0.8	3 5	-3 6	0.4
	18- <i>H</i> ₃	21- <i>H</i> ₃	22	-H	SeCH ₃
13	0.55 / 0 58	1.03 / 0.9°	3 15	/ 2 97	2 06 / 2.02
Appearance ^f	s s	d (6.8) ?	m	bt (7.5)	s s
(22 <i>S/R</i>)-14	0 55 / 0.59	1.03 / 0.9°	3 08	/ 2 98	2 06 / 2.02
Appearance ^f	s s	d (6.9) ?	m	bt (7.5)	s s

a 8 ppm, 300 (¹H) or 75 5 (¹³C) MHz, solvent CDCl₃, ref CHCl₃ = 7 26 or CDCl₃ = 76 8

b Data for 13 (major / minor epimers) are abstracted from the Experimental Section Data for 14 were obtained from the spectra of the available¹³ pure 22S-14 and ca 1 1 mixture of 22S-14 and 22R-14

c Chemical shift difference between major and minor signals

d Chemical shift difference between the signals in 22S-14 and 22R-14

e Approximate position signal partially obscured

f Abbreviations s= singlet, d= doublet, bt= broad triplet, m= multiplet, (Coupling constant J in Hz)

Following the established methodology,¹¹ for the radical deselenation reaction, the conjugated triene molety of 13 was first masked as the cycloaddition products (15) formed with sulphur dioxide (which adds across C-6 and C-19 to give a *ca* $3\cdot 1$ mixture of 6S and 6R adducts) (Scheme 4) Photo-induced reduction with tributyltin hydride then cleanly gave the SO₂-adducts 17 these were separated chromatographically during purification Thermal cheletropic extrusion of SO₂ from either isomer 17 regenerated the 5*E*-vitamin D system in 16, and the synthesis of MC 1090 (8) was completed by the standard sequence of triplet sensitised 5*E* to 5*Z* photo-isomerisation¹⁷ (to 18) followed by desilylation with fluoride



Scheme 4 (Z = $-S_1Me_2Bu'$) 1. SO₂; 11 *n*-Bu₃SnH - hv; separate, 111 NaHCO₃ (boiling EtOH), 1V anthracene - hv; V. *n*-Bu₄N⁺ F⁻.

Incubation of MC 1090 (10 μ g/mL) with Hep 3B cells for 48 h followed by methanol/dichloromethane extraction allowed discernment of both lipid- and aqueous-soluble metabolites.¹⁸ Analysis of the lipid-soluble fraction on HPLC with diode-array detection (for details of the methods, see ref 8) clearly showed

the presence of 5 compounds diplaying the characteristic vitamin D triene chromophore. One of these, formed in substantial amounts (*ca.* 60 ng/10⁷ cells/day), was identified (after isolation) as the C₂₃ alcohol **6** by direct comparison with authentic material¹³ (comigration on two different HPLC systems and mass spectrometry both with and without per-trimethylsilylation) Analysis of the aqueous-soluble fraction from the extraction (for details of the methods, see ref. 2) led to the measurement of large amounts (*ca.* 140 ng/10⁷ cells/day) of calcitroic acid (7) (comigrating on reverse-phase HPLC with authentic material¹³), conclusively identified by isolation and comparison (HPLC and MS) of its methylated (diazomethane) derivative with authentic¹³ methyl calcitroate. The aldehyde **5** was not a detectable metabolite

The identification of both the C_{23} alcohol (6) and calcitroic acid (7) as metabolites of MC 1090 is in accord with our original hypothesis (Scheme 2) on the breakdown of side chain hydroxylated MC 1090, though we have not resolved the question of whether the two metabolites lie on the same or on divergent pathways. If both compounds result from exclusive 25-hydroxylation, then the alcohol is an intermediate, while this is not necessarily the case if 23-hydroxylation is operative. Although further details need clarifying, the results so far have provided the first demonstration that the terminal steps of calcitroic acid production can take place in non-target cells for the vitamin D hormone.

EXPERIMENTAL SECTION

General. Reactions were performed routinely under a nitrogen atmosphere Petroleum ether refers to the pentane fraction Organic solutions were dried over anhyd. MgSO₄ Column chromatography was performed on Merck Kieselgel 60 using the "flash" technique Analytical TLC (to which R_f values refer) was performed on Merck plates pre-coated with silica gel 60 F₂₅₄. Melting points were detemined with a Buchi-Tottoli apparatus and are uncorrected Microanalyses were performed by Messrs G Cornali and W Egger (Leo) UV-Spectra (λ_{max}) were measured for solutions in 96% EtOH on a Perkin Elmer Lambda 5 spectrophotometer ¹H-NMR spectra (δ_H) and ¹³C-NMR spectra (δ_C) (courtesy Mr N R. Andersen, Leo) were run on a Bruker AC-300 spectrometer for solutions in CDCl₃ using the solvent as internal standard (CDCl₃ = 76 8 ppm, residual CHCl₃ = 7 26 ppm these reference values correspond to internal Me₄Si = 0 0(0) ppm), 100 MHz ¹H-NMR spectra (δ_H) were run on a Jeol FX100 70 eV EI mass spectra (m/z) were obtained on a Hewlett-Packard HP5985 mass spectrometer fitted with a direct insertion probe

(15,3R,5E,7E,22Z)-1,3-Bis-(t-butyldimethylsilyloxy)-22-methylselenyl-24-oxa-9,10-secocholesta-5,7,10(19)-triene (13). A sturred solution of the seleno-acetal (11)¹¹ (435 mg, 0 58 mmol) in dry THF (5 mL) was cooled to -70 °C and treated dropwise over 5 min with n-BuLi (1 4 M in hexanes, 0.5 mL, 0 7 mmol) After 10 min, chloromethyl *i*-propyl ether¹⁹ (freshly distilled, 95 mg, 0 87 mmol) was added, and after a further 30 min at -70 °C, the reaction solution allowed to warm to -20 °C over 30 min Ether (30 mL) was then added and the solution was extracted with 2% sodium hydrogen carbonate solution. The organic phase was washed with brine, dried and concentrated in vacuo to give an oil Purification by chromatography (30 g silica gel, 2%, increasing to 5%, ether in petroleum ether as eluant) gave 13 (308 mg, 73%) as an oil that crystallised upon standing The product was chromatographically homogeneous (R_f ca 0 5, 5% ether in petroleum ether as eluant), although the NMR spectra showed that it consisted of a mixture of epimers at C-22 in a 3 2 ratio Recrystallisation from ether-methanol gave an analytical sample (needles, mp 110-112 °C), in which this ratio was increased to ca 2 1, Found C, 64 97, H, 10 04 Calcd for $C_{39}H_{72}O_3SeS_{12}$ C, 64 69; H, 10 02%, λ_{max} 270 nm (ϵ 25500), δ_H (300 MHz) (ca 1[#] 2^{*}/2^{*}.1[#] integrated ratio of appropriate signals from the 22*R*[#] and the 22*S*^{*} isomer respectively) 0 07 (br s, 12 H, SICH3's), 0 55° and 0 58" [2 s (ratio ca 2° 1"), 3 H, 18-H3], 0 87 and 0 90 (each: s, 9 H, SIC(CH3)3), ca 0.9[#] (partially obscured, deduced from C-H correlation) and 1.03^{*} [2 d (ratio ca 1[#] 2^{*}), $J^* = 6.8$ Hz, 3 H, 21-H₃], 1 15, 1.15 and 1 16 (overlapping d's, J = 6 Hz, 6 H, 26- and 27-H₃'s), 2 02[#] and 2 06[•] [2 s (ratio ca 1[#] 2^{*}), 3 H, SeCH₃], 2 30 (br d, J = 14 Hz, 1 H, 4β -H), 2 57 (dd, J = 5, 14 Hz, 1 H, 4α -H),

2 87 (b d, J = 12 Hz, 1 H, 9 β -H), 2.97[#] and 3 15^{*} [b t[#], J = 7.5 Hz, and ddd^{*}, J = 2, 5, 8 Hz (ratio ca. 1[#]:2^{*}), 1 H, 22-H], 3 45-3.79 (m's, 23-H₂ and 25-H), 4 22 (m, 1 H, 3-H), 4.54 (m, 1 H, 1-H), 4.94 (br s, 1 H, 19Z-H), 4.99 (m, 1 H, 19E-H), 5 83 and 6.46 (each: d, J = 11.3 Hz, 1 H, 7-H and 6-H); $\delta_{\rm C}$ (ca. 1[#]:2^{*}/2^{*}:1[#] ratio peak heights where appropriate) -5.1, -5 1, -5.0, -4 9 (S1CH₃'s), 4.1[#], 4.5^{*} (SeCH₃), 11.5^{*}, 12.2[#] (C-18), 14.6[#], 15.6^{*} (C-21), 17.9, 18.1 (S1CMe₃'s), 21 8, 21.9, 22 2 (C-26 and C-27), 22 0 (C-11), 23.3 (C-15), 25.6, 25.7 (S1CCH₃'s), 27 3[#], 27.5^{*} (C-16), 28.7 (C-9), 36.4 (C-4), 36.4[#], 40.3^{*} (C-20), 40.3 (C-12), 43.8 (C-2), 45.6[#], 46.0^{*} (C-23), 71 5[#], 71 8^{*} (C-25), 106.4 (C-19), 116.3[#], 116.4^{*} (C-7), 121.5 (C-6), 135.2[#], 135.4^{*} (C-5), 142.8^{*}, 143 1[#] (C-8) and 153 4 (C-10)

(1*S*,3*R*,5*E*,7*E*)-1,3-Bis-(*t*-butyldimethylsilyloxy)-24-oxa-9,10-secocholesta-5,7,10(19)-triene (6R)and (6S)-sulphur dioxide adducts (17). Compound 13 (mixture of diastereoisomers, 360 mg, 0.50 mmol) was dissolved in ether (0 5 mL) and liquid SO₂ (10 mL) added The SO₂ was allowed to boil under reflux for 30 min and then the solvents were removed in vacuo to give a diastereoisomeric mixture of SO₂adducts of the compounds 13, showing two spots on TLC (R's ca 05 and 0.3, 40% ether in petroleum ether as eluant). [The ¹H-NMR spectra of the two components, separated in a ca 3:1 ratio from the product of a parallel experiment, showed the major (less polar) to be the 22-epimeric mixture of (6S)sulphur dioxide adducts (6S)-15 (18-H₃ singlets at δ 0.66^{*} and 0.69^{*}), while the minor component was the 22-epimeric mixture of (6R)-sulphur dioxide adducts (6R)-15 (18-H₃ singlets at δ 0.56^{*} and 0.59[#]) (cf ref. 11)] The product (in a pyrex flask) was dissolved in toluene (14 mL) and n-Bu₃SnH (0.87 g, 3 mmol) added. A nitrogen atmosphere was established, and the reaction flask was water cooled (20 °C) during illumination with radiation from a high pressure Hg lamp (type, Hanau TO 718Z2) for 75 min The solution was then concentrated in vacuo to give a residue containing the title compounds, which was purified by chromatography (50 g silica gel, 30% ether in petroleum ether as eluant). First eluted was the major isomer (6S)-17 (243 mg, 70%), needles from ether-methanol, mp 124-125 °C; Found: C, 65.63; H, 10 21; S, 4 50 Calcd for C₃₈H₇₀O₅SS1₂ C, 65 65, H, 10 15, S, 4 61%, $\delta_{\rm H}$ (100 MHz) 0 06 (br s, 12 H, S_1CH_3 's), 0 65 (s, 3 H, 18-H₃), 0 87 and 0 88 (each s, 9 H, $S_1C(CH_3)_3$), 0.94 (d, J = 6 Hz, 3 H, 21-H₃), 1 13 (d, J = 6 Hz, 6 H, 26- and 27-H₃'s), 3.2-4.1 (m's, including 3 53, hept, J = 6 Hz, 25-H, and 3.76, br ABq, J = 16 Hz, 19-H₂, 5 H, including also 23-H₂), 4 2 (m, 1 H, 3-H), 4 35 (m, 1 H, 1-H), 4.67 (m, 2 H, 6-H and 7-H) This was followed by the minor isomer (6R)-17 (76 mg, 22%), obtained as a gum, $\delta_{\rm H}$ (100 MHz) 0 07 (br s, 12 H, SiCH₃'s), 0 58 (s, 3 H, $18-H_3$), 0 89 and 0 90 (each s, 9 H, SiC(CH₃)₃), 0.95 (d, J = 7 Hz, 3 H, 21-H₃), 1 15 (d, J = 6 Hz, 6 H, 26- and 27-H₃'s), 3 25 - 4 1 (m's, including 3 53, hept, J = 6 Hz, 25-H, and 3 77, br ABq, J = 16 Hz, 19-H₂, 5 H, also including 23-H₂), 4.15 (m, 1 H, 3-H), 4 4 (m, 1 H, 1-H), 4 5-4 95 (2 br d, 2 H, J = 10 Hz, 6-H and 7-H)

(15,3*R*,5*E*,7*E*)-1,3-Bis-(*t*-butyldimethylsilyloxy)-24-oxa-9,10-secocholesta-5,7,10(19)-triene (16). Compound [17, major (6S) isomer] (183 mg, 0.26 mmol) together with NaHCO₃ (0.3 g) was dissolved or suspended in 96% ethanol (10 mL), and the stirred mixture was heated under reflux for 90 min. After cooling, the reaction mixture was partitioned between EtOAc (30 mL) and water, and the EtOAc layer was washed with brine and dried Removal of the solvent *in vacuo* and purification by chromatography (15 g silica gel, 3% ether in petroleum ether as eluant) followed by crystallisation from ether-methanol gave 16 (140 mg, 84%) as needles, mp 76-77 °C, Found C, 72.39, H, 11.18 Calcd for C₃₈H₇₀O₃SSl₂· C, 72.32; H, 11 18%; λ_{max} 270 nm; $\delta_{\rm H}$ (100 MHz) 0 07 (br s, 12 H, SiCH₃'s), 0.56 (s, 3 H, 18-H₃), 0.88 and 0.91 (each s, 9 H, SiC(CH₃)₃), 0.95 (d, J = 7 Hz, 3 H, 21-H₃), 1.15 (d, J = 6 Hz, 6 H, 26- and 27-H₃'s), 3 2-3 7 (m, including 3 55, hept, J = 6 Hz, 25-H, 3 H, including 23-H₂), 4.22 (m, 1 H, 3-H), 4.54 (dd, J = 5, 9 Hz, 1 H, 1-H), 4.97 (m, 2 H, 19-H₂), 5.82 and 6.47 (each d, J = 11 Hz, 1 H, 7-H and 6-H)

(15,3R,5Z,7E)-24-Oxa-9,10-secocholesta-5,7,10(19)-triene-1,3-diol (MC 1090, 8). A solution of compound (16) (49 mg, 0.08 mmol), anthracene (25 mg) and triethylamine (0.1 mL) in toluene (5 mL) in a pyrex flask was irradiated with light from a high pressure ultraviolet lamp, type TQ150Z2 (Hanau) at about 20°C for 1 h The reaction mixture was filtered, concentrated *in vacuo* and purified by chromatography (15 g silica gel, 2% ether in petroleum ether as eluant) to give (15,3R,5Z,7E)-1,3-bis-(*t*-

butyldimethylsilyloxy)-24-oxa-9,10-secocholesta-5,7,10(19)-triene (18) (44 mg), obtained as a gum [λ_{max} 265 nm; δ_{H} (100 MHz) 0.07 (br s, 12 H, S1CH₃'s), 0.54 (s, 3 H, 18-H₃), 0.88 (br s, 18 H, S1C(CH₃)₃'s), 0.95 (d, J = 6.5 Hz, 3 H, 21-H₃), 1 15 (d, J = 6 1 Hz, 6 H, 26- and 27-H₃'s), 3.25-3.70 (m, including 3.55, hept, J = 6.1 Hz, 25-H, 3 H, including 23-H₂), 4.19 (m, 1 H, 3-H), 4.37 (m, 1 H, 1-H), 4 87 (d, J = 3 Hz, 1 H, 19E-H), 5.18 (m, 1 H, 19Z-H), 6.01 and 6.23 (each d, J = 11 Hz, 1 H, 7-H and 6-H).] A solution of the entire product (0.07 mmol) and n-Bu₂N⁺ F⁻ (trihydrate, 0.1 g, 0.32 mmol) in THF (5 mL) was stirred at 60 °C for 50 min. The reaction mixture was partitioned between ethyl acetate (30 mL) and 2% sodium hydrogen carbonate solution (20 mL). The organic phase was washed with brine, dried and concentrated in vacuo to give an oil Purification by chromatography (15 g silica gel, 66% ethyl acetate in petroleum ether as eluant) gave the title compound 8 (21 mg, 67% from 16), obtained as a gum; λ_{max} 264 nm (ϵ 17500); $\delta_{\rm H}$ (300 MHz) 0.55 (s, 3 H, 18-H₃), 0.95 (d, J = 6.5 Hz, 3 H, 21-H₃), 1 15 (d, J = 6.0Hz, 6 H, 26- and 27-H₃'s), 2.31 (dd, J = 7, 13 Hz, 1 H, 4β -H), 2 59 (dd, J = 3, 13 Hz, 1 H, 4α -H), 2.82 (dd, J = 3, 11 Hz, 1 H, 9β -H), 3 33-3.60 (m, including 3.54, hept, J = 6.1 Hz, 25-H, 3 H, ncluding 23-H2), 4.23 (m, 1 H, 3-H), 4 43 (m, 1 H, 1-H), 5.00 (br s, 1 H, 19E-H), 5.33 (br s, 1 H, 19Z-H), 6.02 and 6.38 (each d, J = 11.3 Hz, 1 H, 7-H and 6-H), δ_{c} 12 0, 19.2, 22.1, 22.3, 22.3, 23.6, 27.6, 29.1, 33.7, 36.1, 40 5, 42 9, 45 3, 46.0, 56 4, 56.9, 66 3, 66.9, 70.9, 71 3, 111.8, 117.1, 125.0, 132.9, 143.2, and 147.6, m/z 402 (M⁺, 6), 387 (0 5), 384 (6), 369 (1), 366 (3), 343 (1), 287 (2), 269 (3), 251 (5), 152 (30), and 134 (100%), HRMS (EI) calcd for C₂₆H₄₂O₃ 402 3134 (M⁺); found: 402.3139.⁹

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